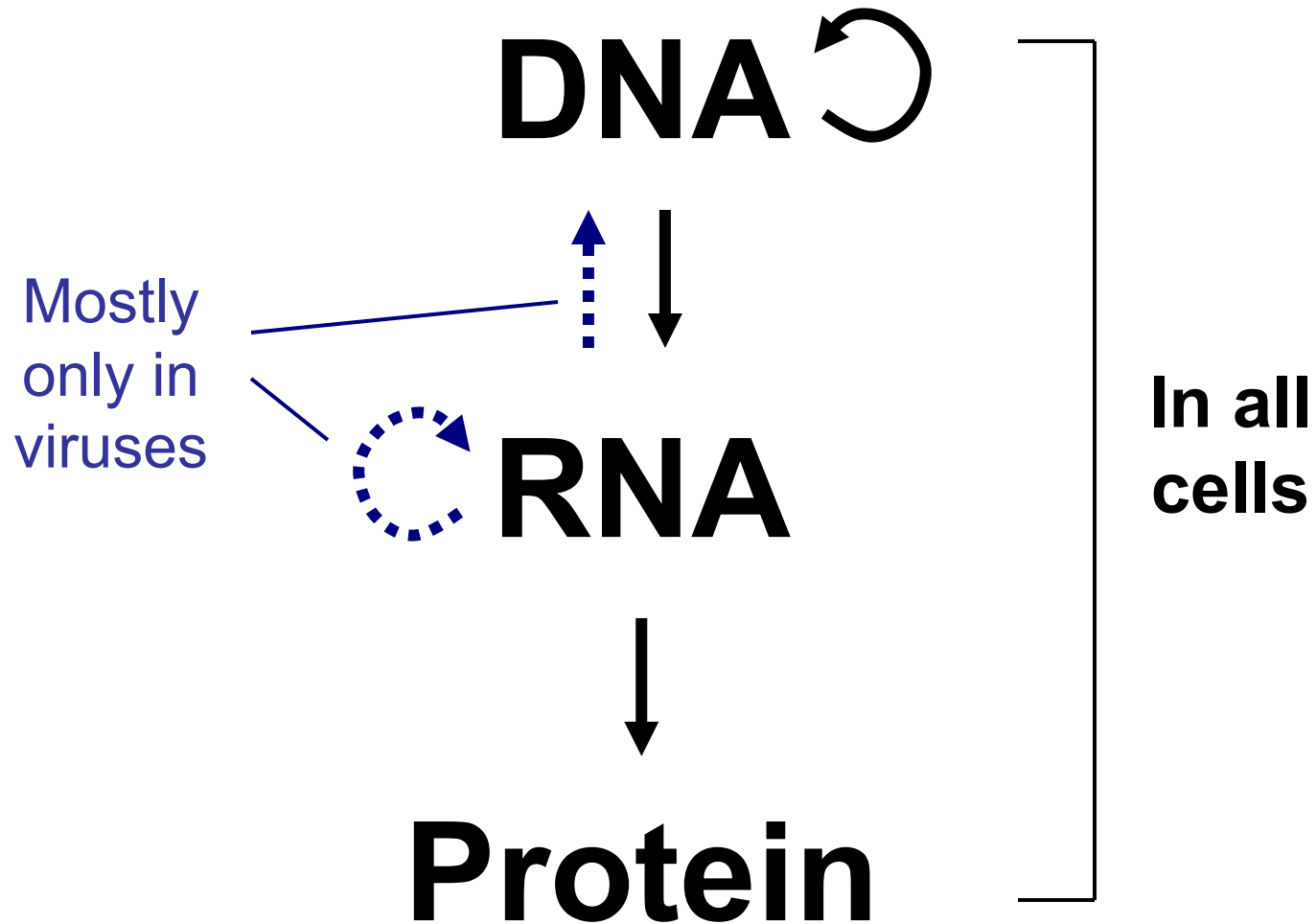
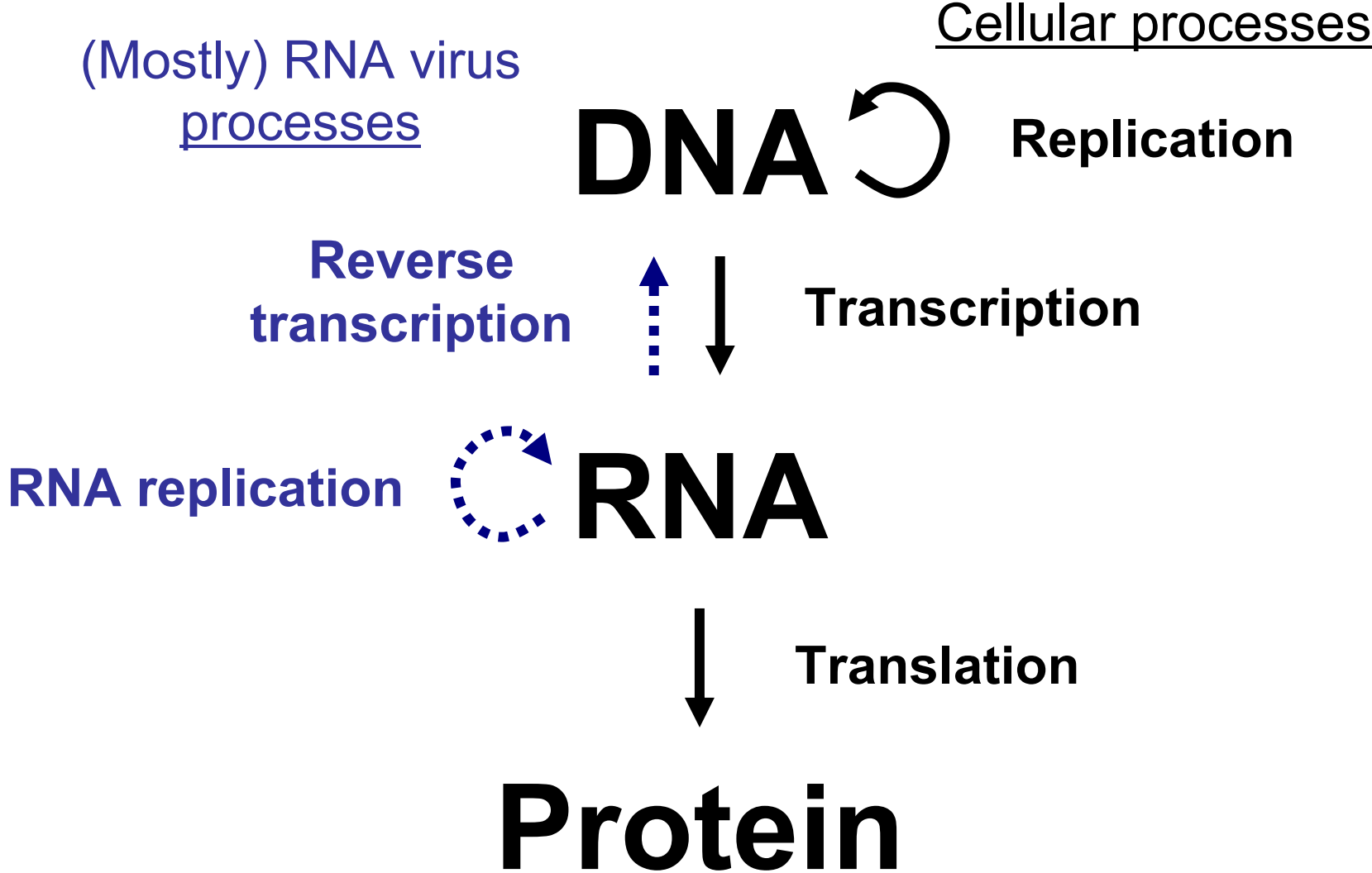


Central Dogma of Biology



Processes in the central dogma



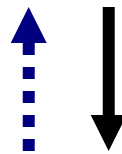
Enzymes in the central dogma

Cellular enzymes

(Mostly) RNA virus
enzymes

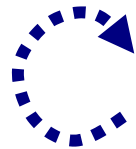
DNA  **DNA polymerase**

**Reverse
transcriptase**



RNA polymerase

**RNA-dependent
RNA polymerase**



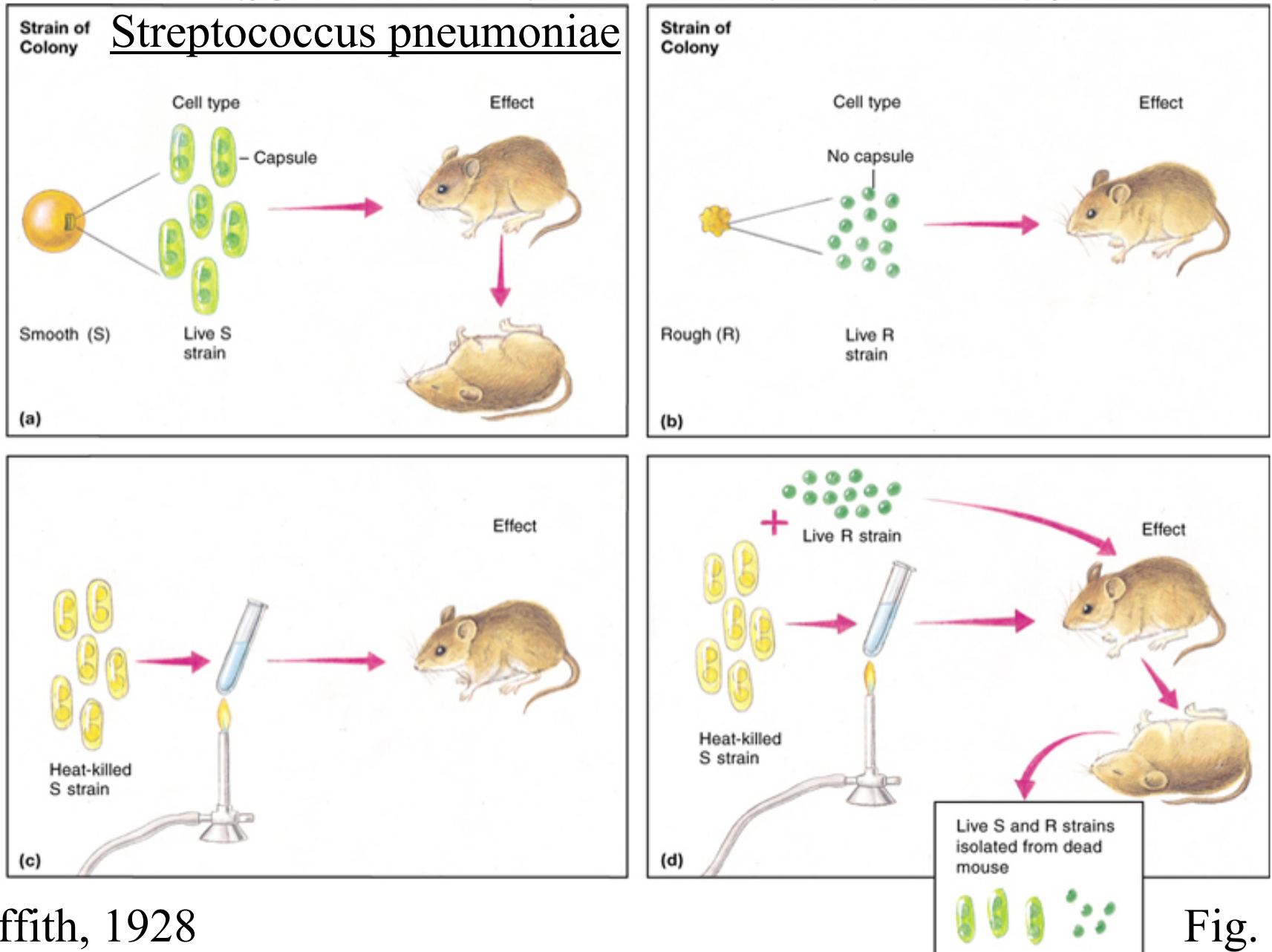
RNA



Ribosome

Protein

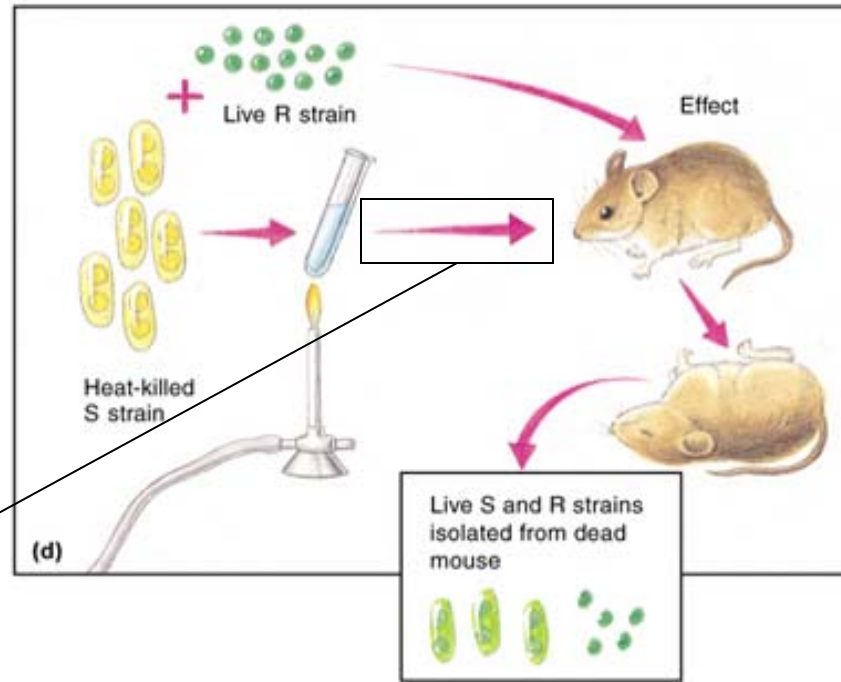
Genetic material is transferable between bacteria



Griffith, 1928

Fig. 2.2

The genetic material is DNA



Heat killed S strain treated with:

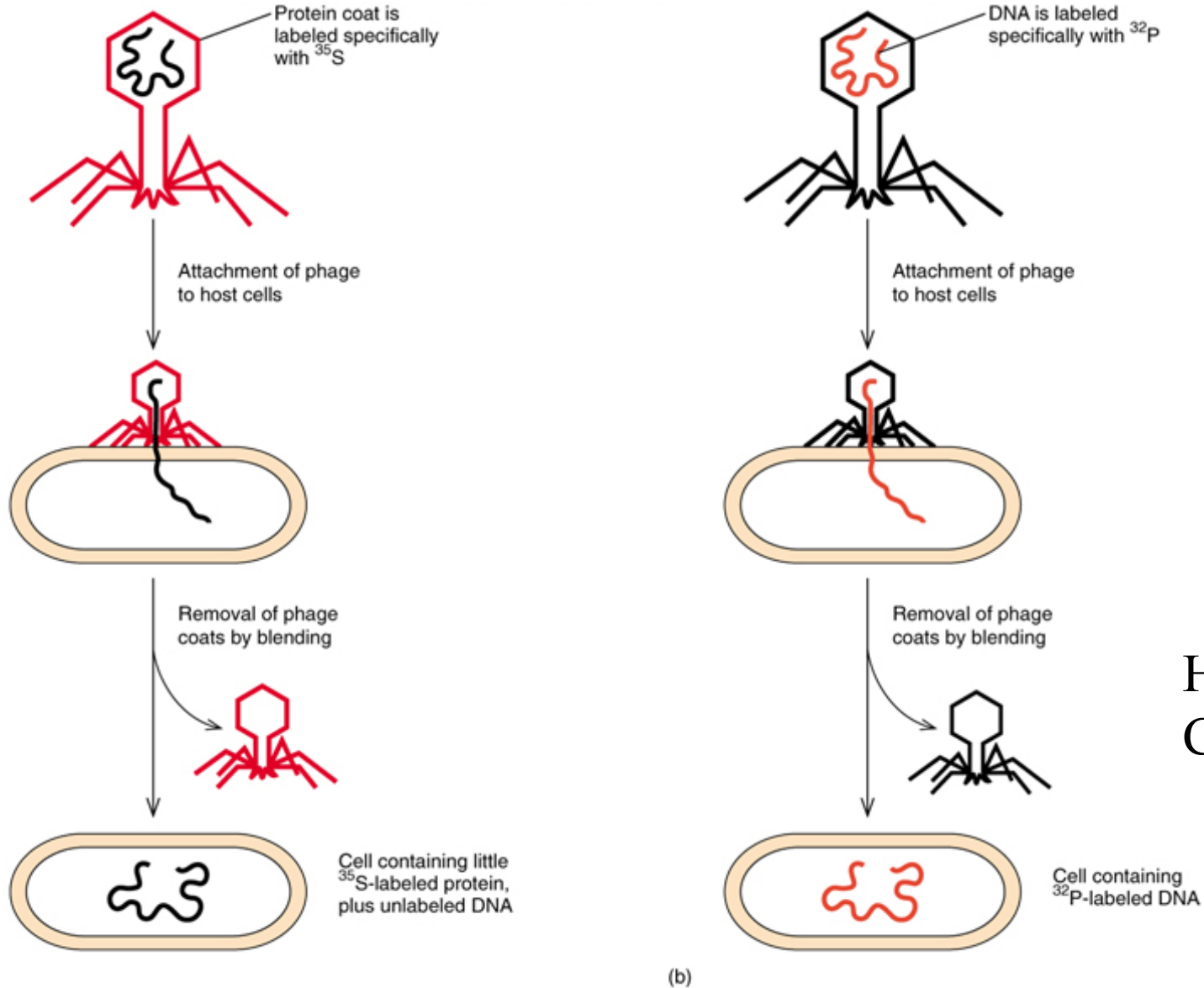
- Phenol => still infectious
- Proteinase (trypsin, chymotrypsin) => still infectious
- UV light => no longer infectious
- RNase => still infectious
- DNase => no longer infectious

=> Genetic material is DNA!

Avery, MacLeod,
McCarthy, 1944

Based on Fig. 2.2

The genetic material of bacteriophage T2 is DNA



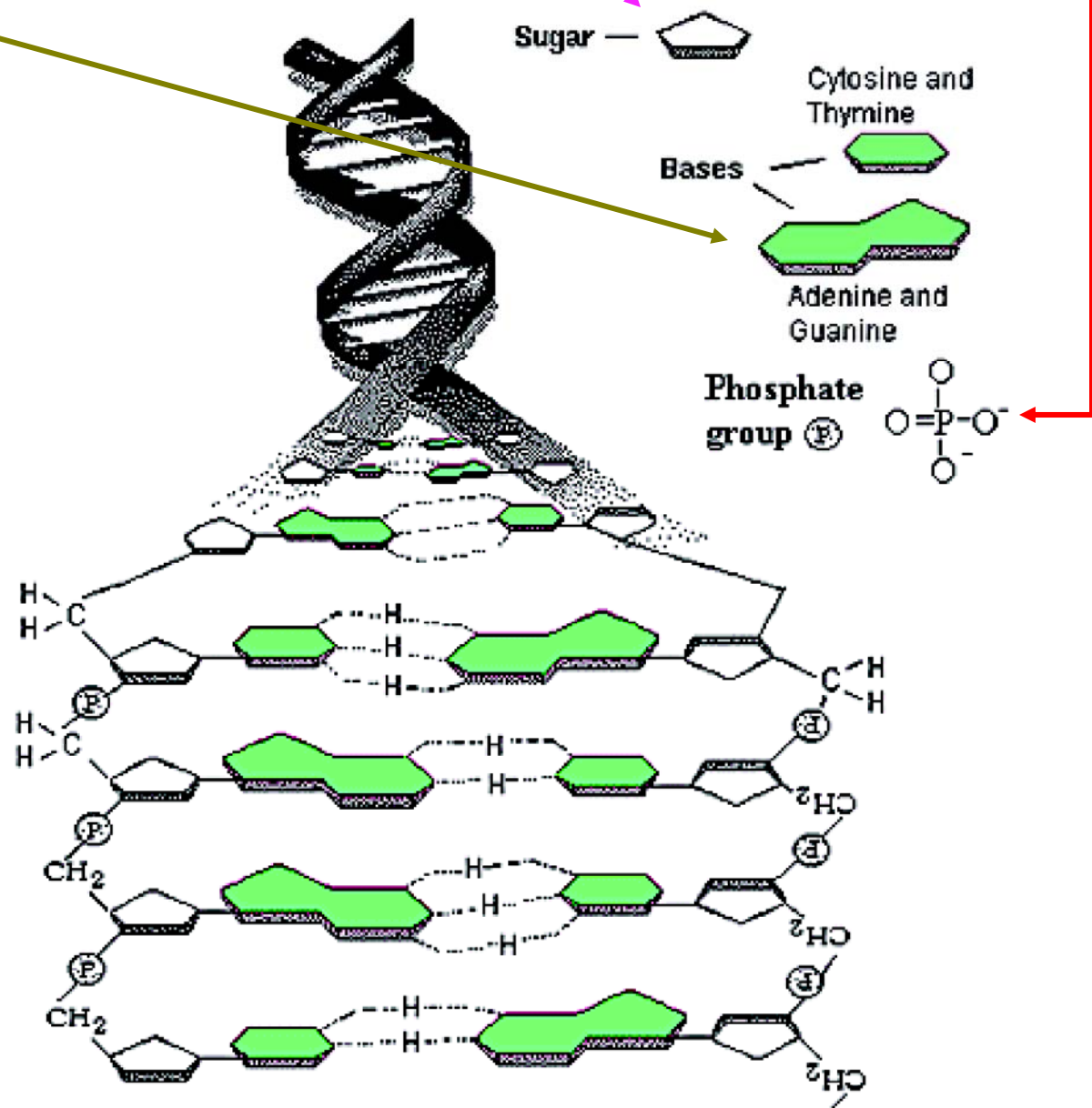
Hershey,
Chase, 1952

Fig. 2.4

Some viruses use RNA as genetic material (e.g. HIV, herpesviruses etc)!

- Clicker Question -

A nucleotide is composed of a **sugar**, a **phosphate** and a **base**



The chemical nature of DNA and RNA

DNA

Nitrogenous bases:

Adenine (A)

Cytosine (C)

Guanine (G)

Thymine (T)

Phosphoric acid

Sugar: 2' deoxyribose

RNA

Nitrogenous bases:

Adenine (A)

Cytosine (C)

Guanine (G)

Uracil (U)

Phosphoric acid

Sugar: ribose

The sugars

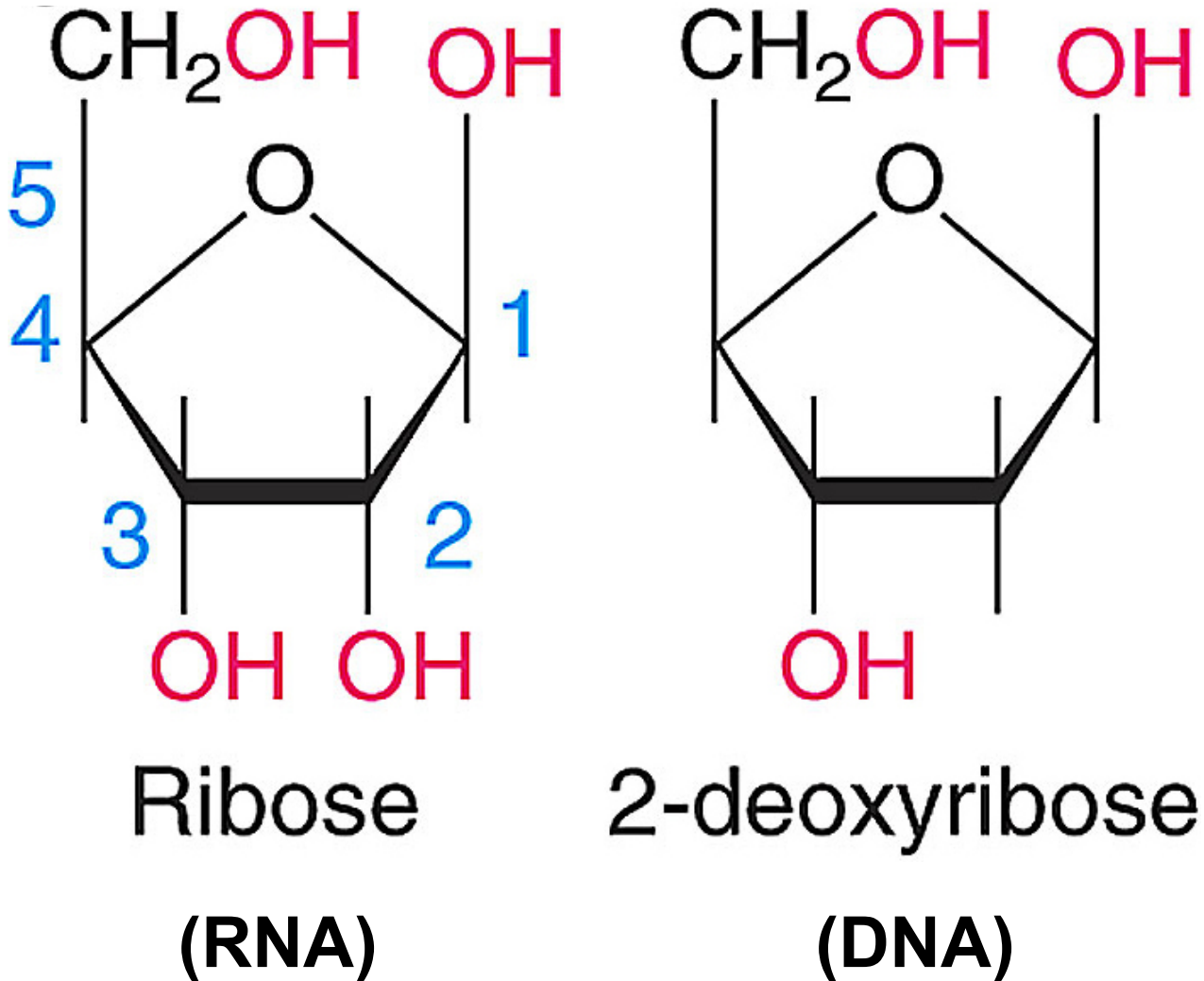


Fig. 2.6

The bases

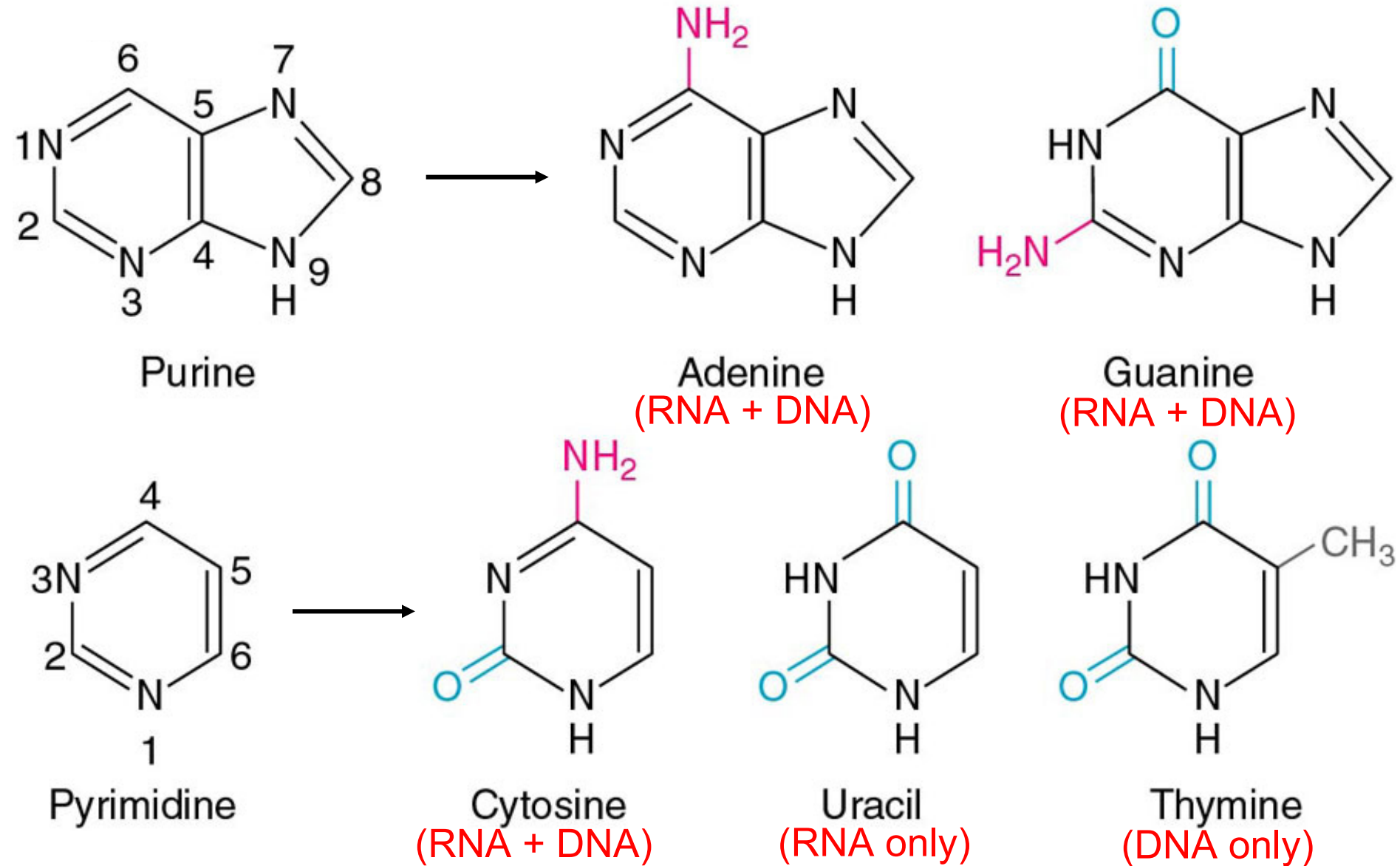


Fig. 2.5

Nucleosides (=sugar+base, no phosphate)

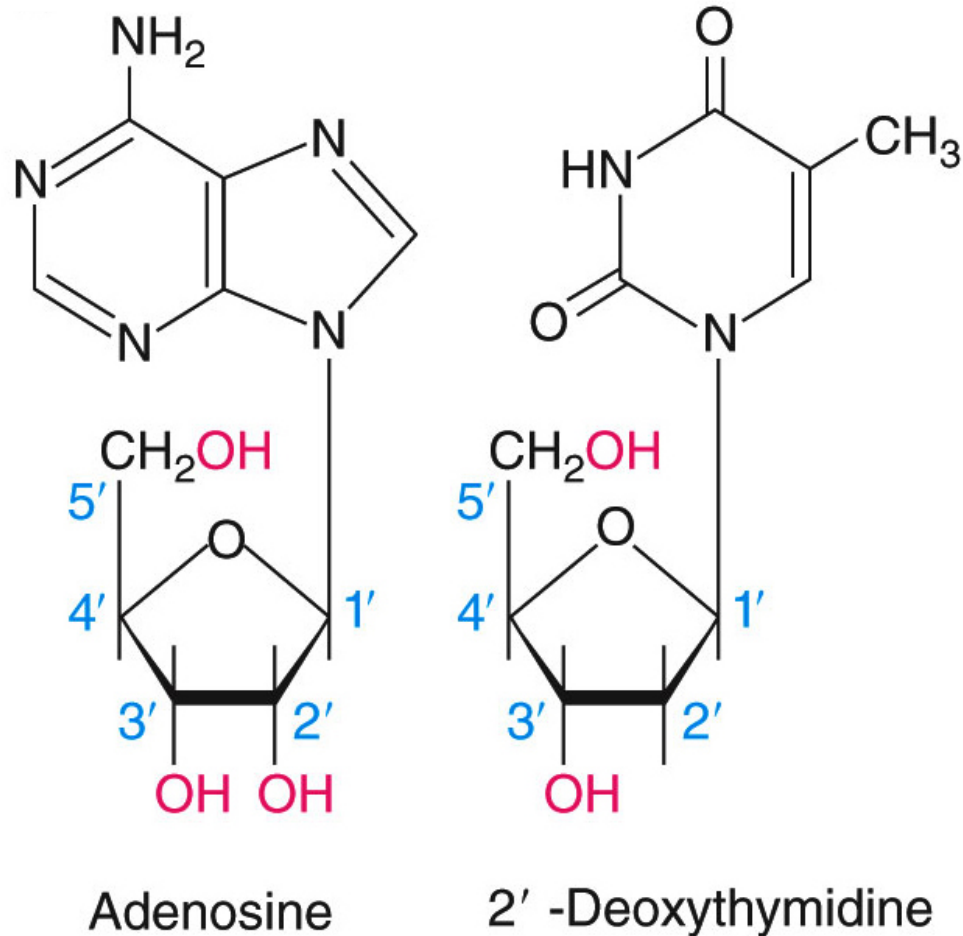
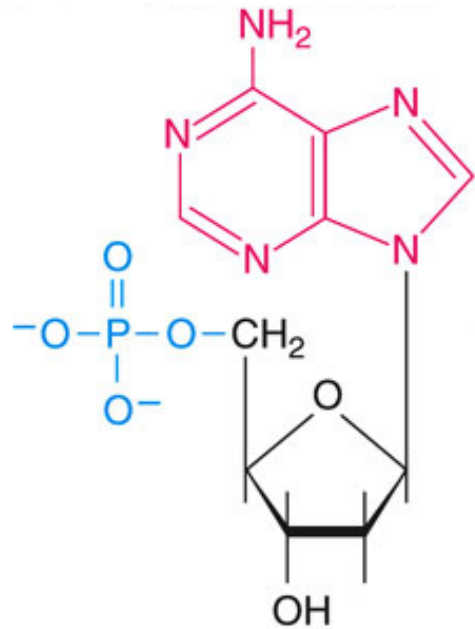
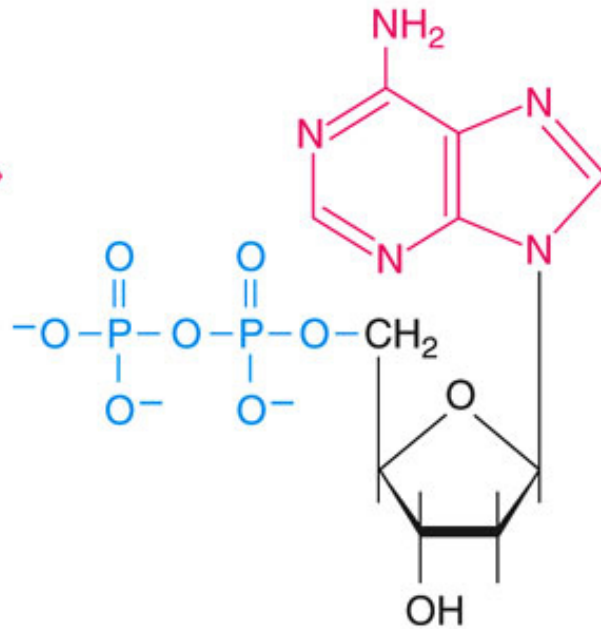


Fig. 2.9

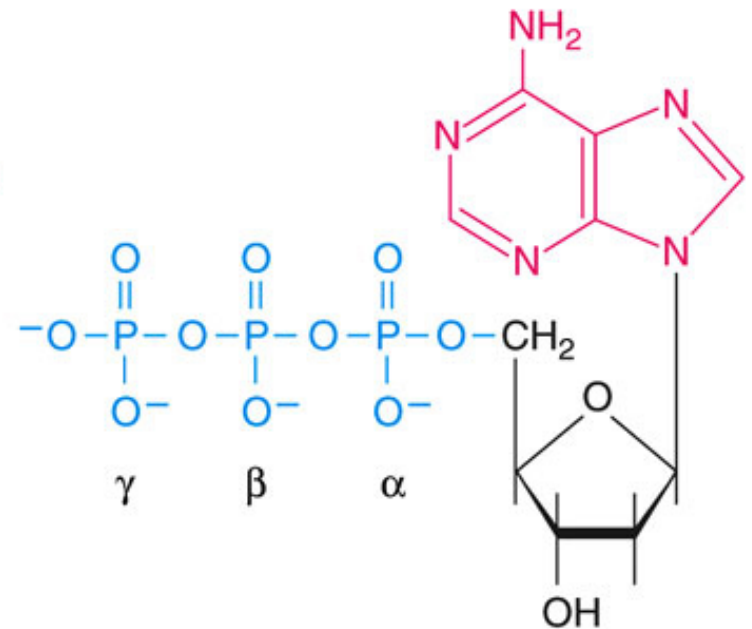
Nucleotides (= nucleoside phosphates)



Deoxyadenosine-5'-
monophosphate (dAMP)



Deoxyadenosine-5'-
diphosphate (dADP)



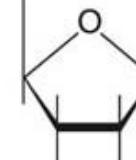
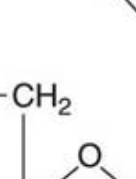
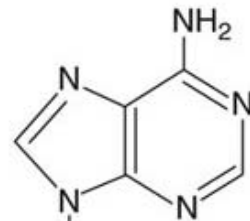
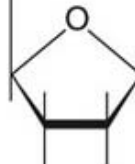
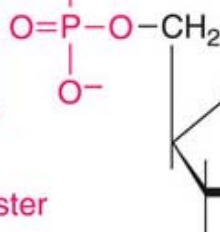
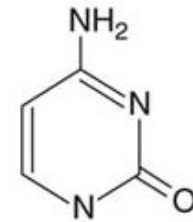
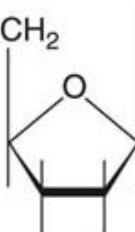
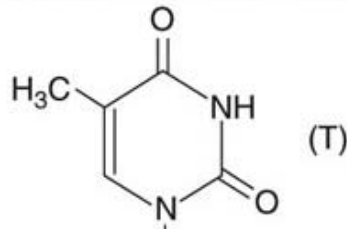
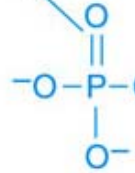
Deoxyadenosine-5'-
triphosphate (dATP)

- Clicker Question -

Nucleotide polymer

5' end

5'-phosphate



Phosphodiester bonds

5' TCA 3'

Figure 2.10

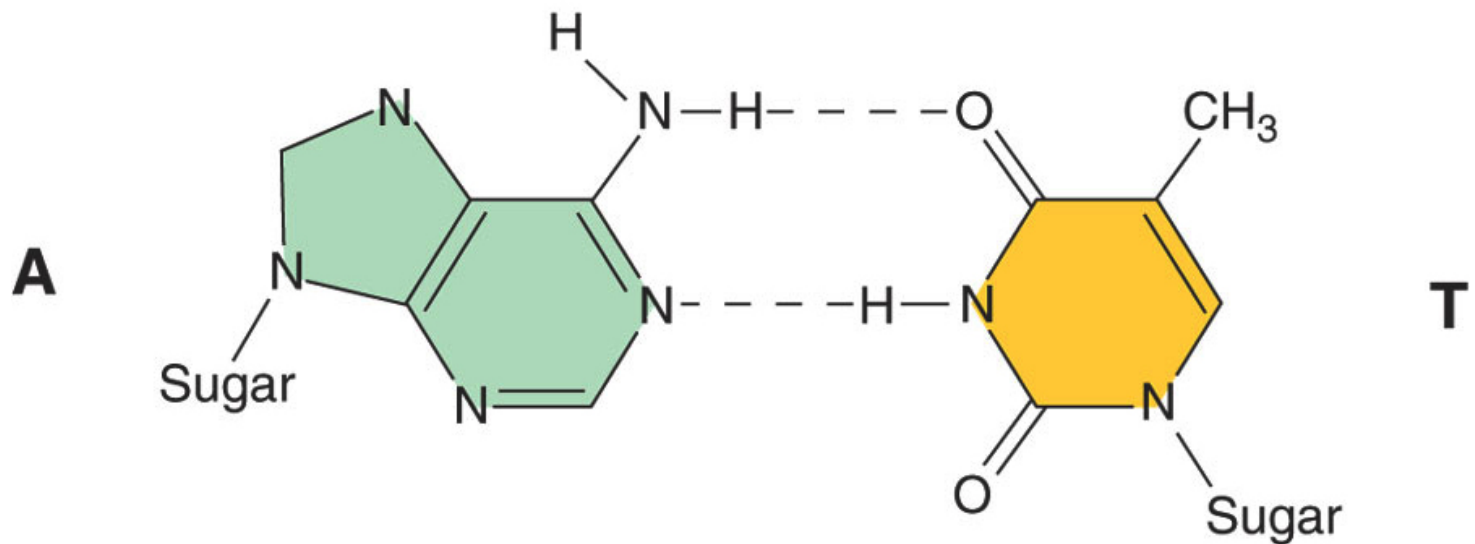
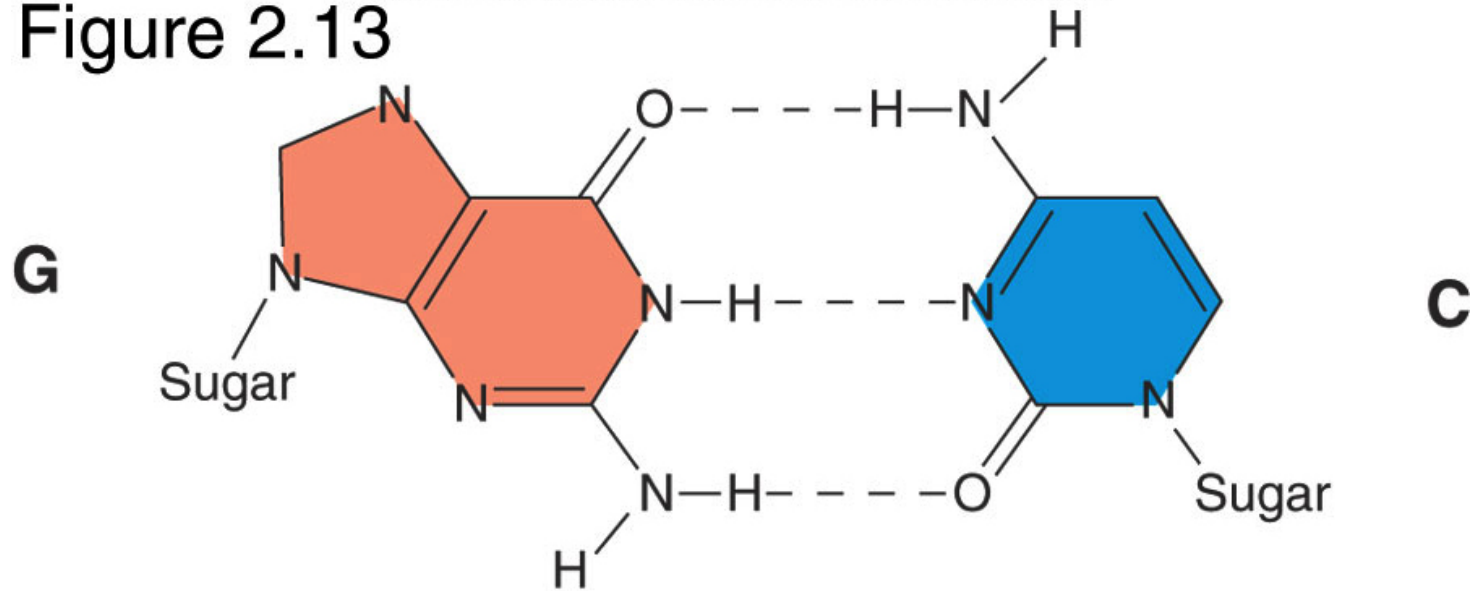
3'-hydroxyl



3' end

Base pairs

Figure 2.13



The double-helix

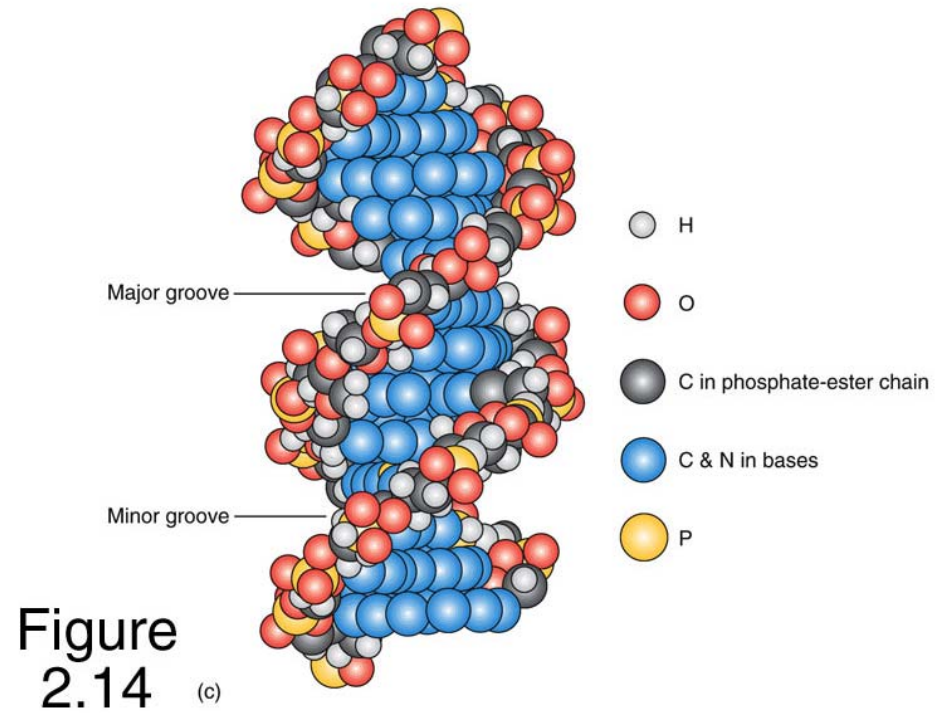
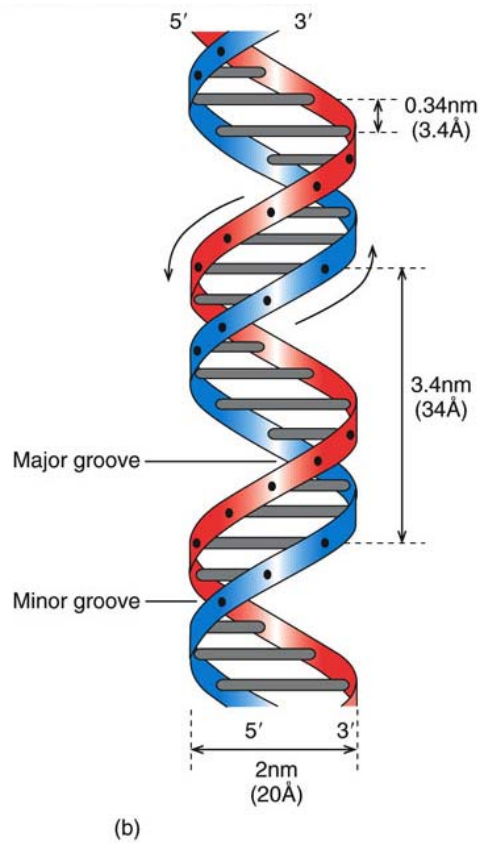
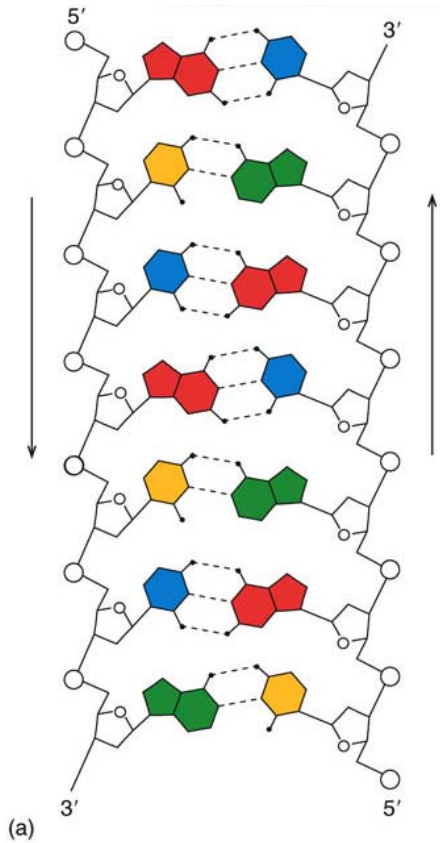
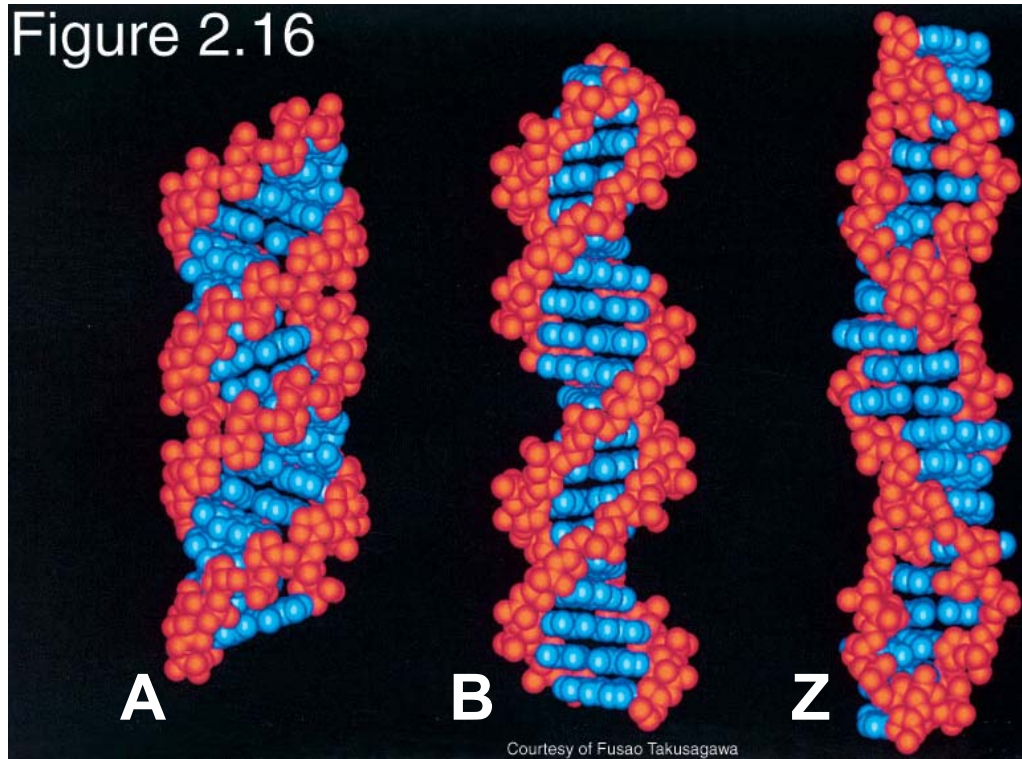


Figure 2.14 (c)

Different polynucleotide double-helix structures

Figure 2.16



Courtesy of Fusao Takusagawa

| | <u>Form</u> | <u>Pitch (Å)</u> | <u>Residues per turn</u> | <u>Inclination of base pair from horizontal</u> |
|---------------------|-------------|------------------|--------------------------|---|
| Most dsRNA → | A | 24.6 | ~11 | +19° |
| Most DNA → | B | 33.2 | ~10 | -1.2° |
| Rare → | Z | 45.6 | 12 | -9° |

- Clicker Question -

Although G=C and A=T are true for every organism, the ratio of G+C versus A+T vary from organism to organism

Table 2.3 Relative G + C Contents of Various DNAs

| Sources of DNA | Percent (G + C) |
|-----------------------------------|-----------------|
| <i>Dictyostelium</i> (slime mold) | 22 |
| <i>Streptococcus pyogenes</i> | 34 |
| Vaccinia virus | 36 |
| <i>Bacillus cereus</i> | 37 |
| <i>B. megaterium</i> | 38 |
| <i>Hemophilus influenzae</i> | 39 |
| <i>Saccharomyces cerevisiae</i> | 39 |
| Calf thymus | 40 |
| Rat liver | 40 |
| Bull sperm | 41 |
| <i>Streptococcus pneumoniae</i> | 42 |
| Wheat germ | 43 |
| Chicken liver | 43 |
| Mouse spleen | 44 |
| Salmon sperm | 44 |
| <i>B. subtilis</i> | 44 |
| T1 bacteriophage | 46 |
| <i>Escherichia coli</i> | 51 |
| T7 bacteriophage | 51 |
| T3 bacteriophage | 53 |
| <i>Neurospora crassa</i> | 54 |
| <i>Pseudomonas aeruginosa</i> | 68 |
| <i>Sarcina lutea</i> | 72 |
| <i>Micrococcus lysodeikticus</i> | 72 |
| Herpes simplex virus | 72 |
| <i>Mycobacterium phlei</i> | 73 |

The DNA content varies between organisms

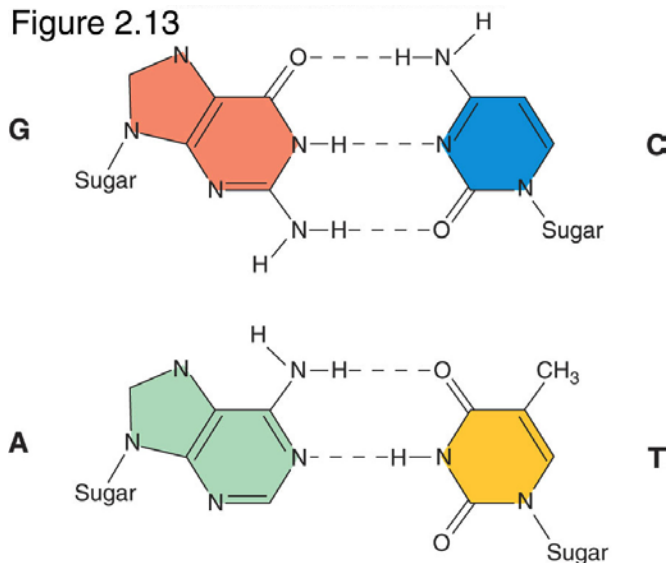
Figure T2.4

Table 2.4 Sizes of various DNAs

| Source | Molecular weight | Base pairs | Length |
|--|---------------------------|---------------------------|-------------------|
| Subcellular Genetic Systems: | | | |
| SV40 (mammalian tumor virus) | 3.5×10^6 | 5226 | 1.7 μm |
| Bacteriophage ϕX174 (double-stranded form) | 3.2×10^6 | 5386 | 1.8 μm |
| Bacteriophage λ | 3.3×10^6 | 5×10^4 | 13 μm |
| Bacteriophage T2 or T4 | 1.3×10^8 | 2×10^5 | 50 μm |
| Human mitochondria * | 9.5×10^6 | 16,596 | 5 μm |
| Prokaryotes: | | | |
| <i>Hemophilus influenzae</i> * | 1.2×10^9 | 1.83×10^6 | 620 μm |
| <i>Escherichia coli</i> | 3.1×10^9 | 4.65×10^6 | 1.6 mm |
| <i>Salmonella typhimurium</i> | 8×10^9 | 1.1×10^7 | 3.8 mm |
| Eukaryotes (content per haploid nucleus): | | | |
| <i>Saccharomyces cerevisiae</i> (yeast) | 7.9×10^9 | 1.2×10^7 | 4.1 mm |
| <i>Neurospora crassa</i> (pink bread mold) | $\sim 1.9 \times 10^{10}$ | $\sim 2.7 \times 10^7$ | ~ 9.2 mm |
| <i>Drosophila melanogaster</i> (fruit fly) | $\sim 1.2 \times 10^{11}$ | $\sim 1.8 \times 10^8$ | ~ 6.0 cm |
| <i>Rana pipiens</i> (frog) * | $\sim 1.4 \times 10^{13}$ | $\sim 2.3 \times 10^{10}$ | ~ 7.7 m |
| <i>Mus musculus</i> (mouse) | $\sim 1.5 \times 10^{12}$ | $\sim 2.2 \times 10^9$ | ~ 75 cm |
| <i>Homo sapiens</i> (human) * | $\sim 2.3 \times 10^{12}$ | $\sim 3.5 \times 10^9$ | ~ 120 cm |
| <i>Zea mays</i> (corn, or maize) | $\sim 4.4 \times 10^{12}$ | $\sim 6.6 \times 10^9$ | ~ 2.2 m |
| <i>Lilium longiflorum</i> (lily) * | $\sim 2 \times 10^{14}$ | $\sim 3 \times 10^{11}$ | ~ 100 m |

Two DNA strands can be separated by heating, a process called DNA denaturation or DNA melting

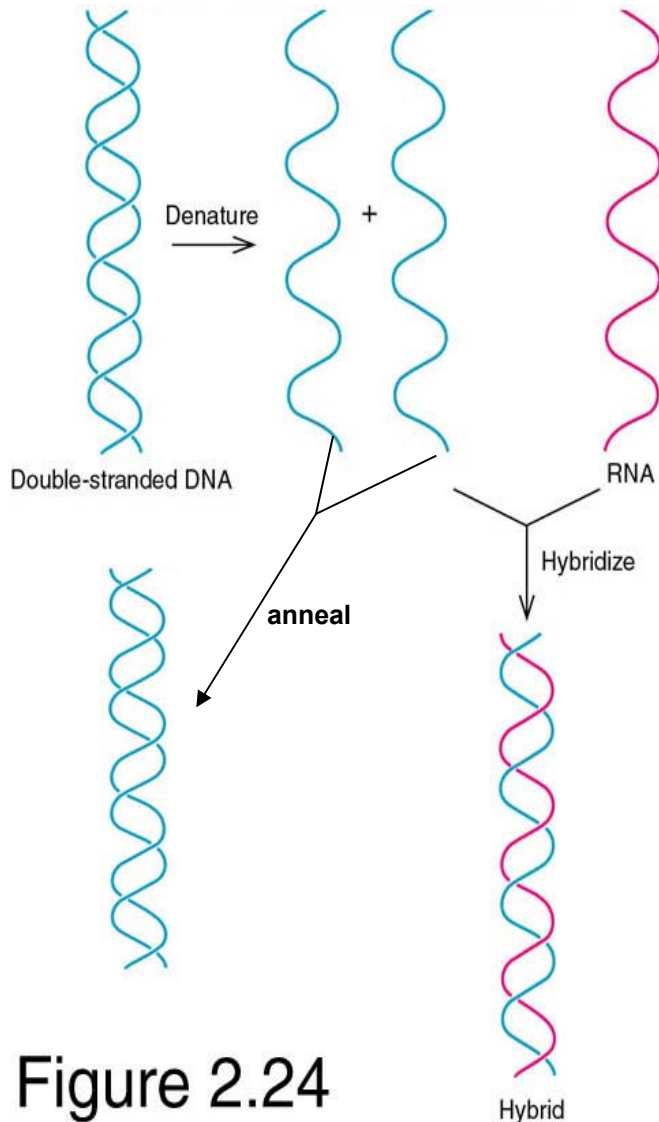
Figure 2.13



The temperature at which the DNA strands are half denatured is called the melting temperature, or T_m .

T_m of a DNA is largely determined by its G/C% (The more G/C the higher T_m) and the length (the longer the higher T_m) (Think of it as the more total hydrogen bonds, the higher temp to denature).

Other ways of denaturing DNA?



Annealing: The process of reuniting separated DNA strands (also called renaturation).

Hybridization: The process of annealing a DNA strand with a complementary RNA strand or DNA strand from a different origin.

Figure 2.24

- Clicker Question -