

# **Lab Math**

## **Primer Preparation and Dilution**

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**March 11, 2015**  
**APHL-CDC**

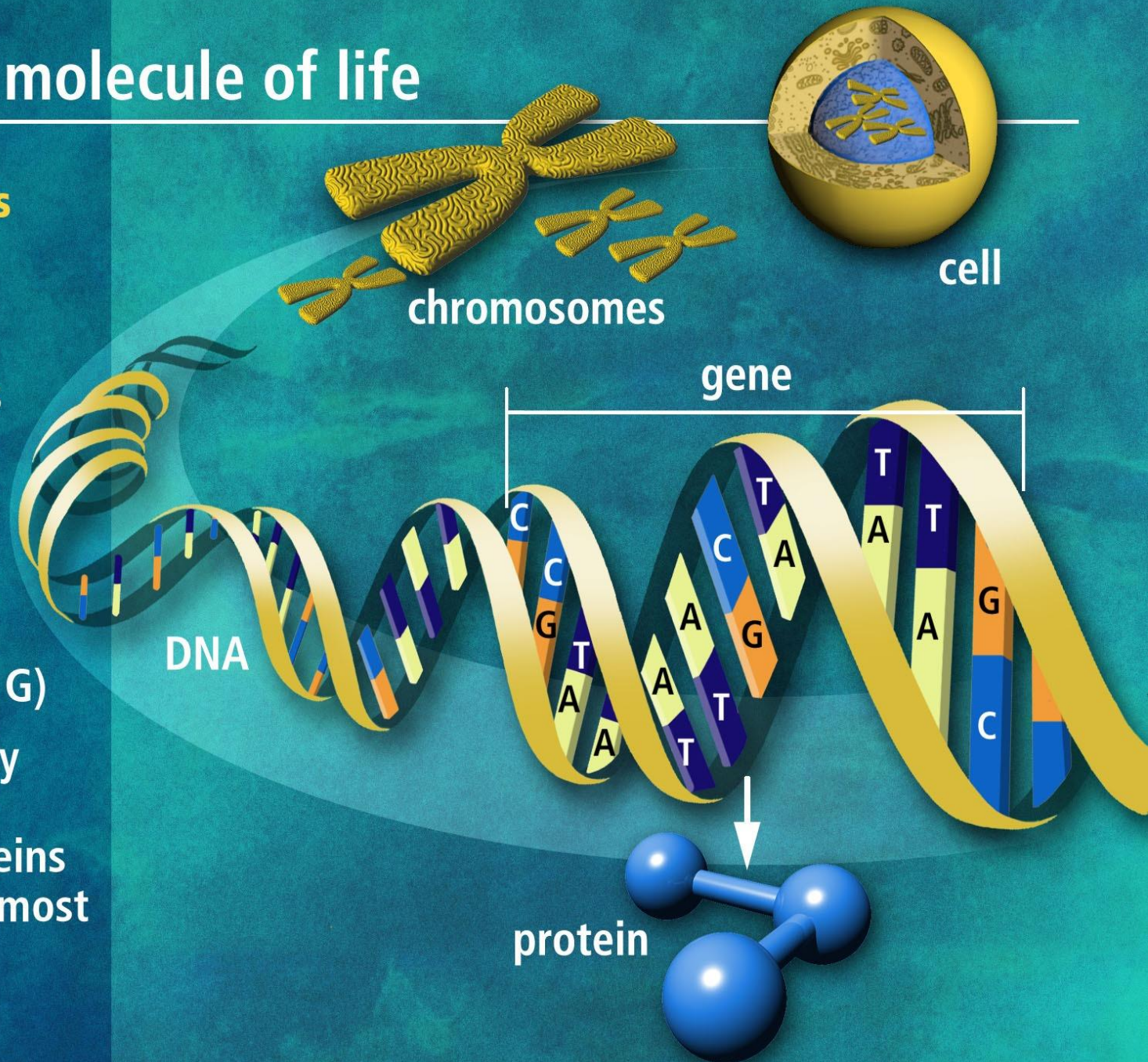


# DNA the molecule of life

## Trillions of cells

Each cell:

- 46 human chromosomes
- 2 meters of DNA
- 3 billion DNA subunits (the bases: A, T, C, G)
- Approximately 30,000 genes code for proteins that perform most life functions





## PCR Cycle – Step 3

**Taq Polymerase Catalyzes  
Primer Extension As  
Complementary Nucleotides  
Are Incorporated**

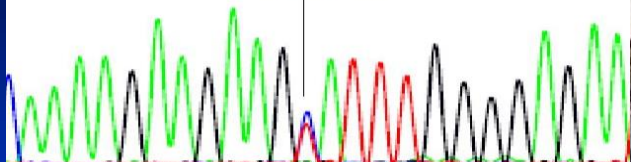


AMT



CAAAAGAAAGAAAGTATTTGGGGAGAA

p.Y146H = c.433T>C



## Concentration of a Solution:

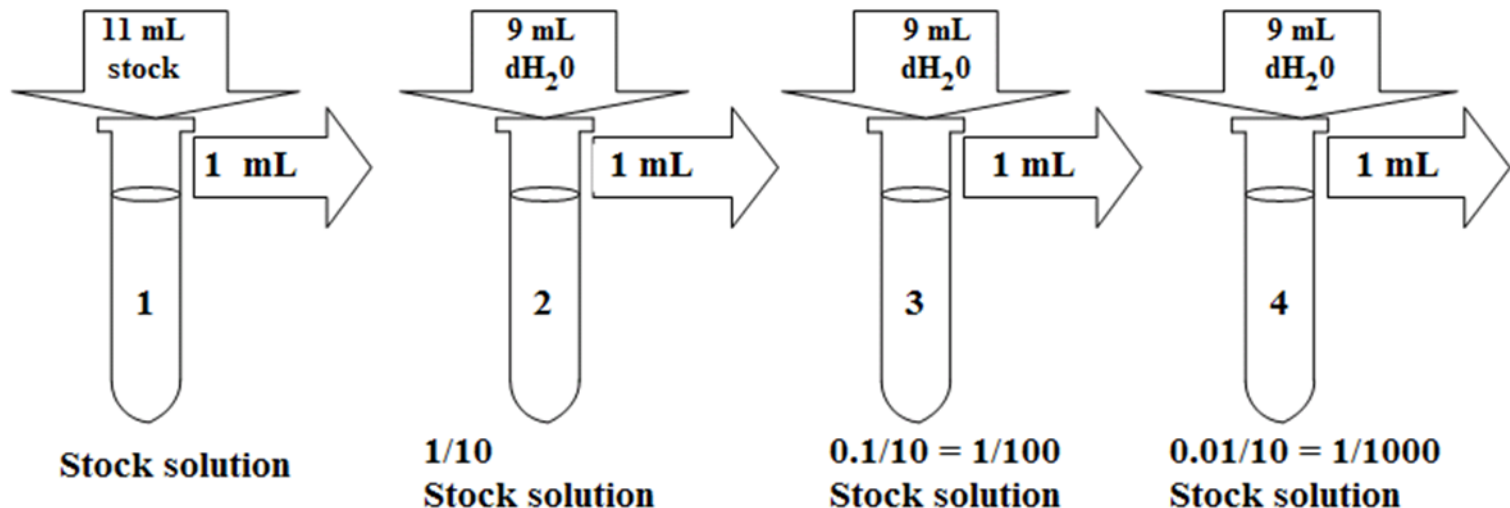
**1M** solution of **NaOH** ( $^{23}\text{Na}^{16}\text{O}^1\text{H}$ ) = 40 g NaOH dissolved in 1 liter of  $\text{H}_2\text{O}$  = **1N**

**1mM** (miliM) = 1:1000 dilution of 1M or  $10^{-3}$

**1 $\mu$ M** (microM) = 1:1000 dilution of 1mM or 1:1,000,000 of 1M or  $10^{-6}$

**1nM** (nanoM) = 1:1000 dilution of 1  $\mu$ M or 1:1,000,000,000 of 1M or  $10^{-9}$

**1pM** (picoM) = 1:1000 dilution of 1 nM or 1:1,000,000,000,000 of 1M or  $10^{-12}$



**Demonstration:  $100 \mu\text{M} = 100 \mu\text{Moles/L} = 0.1 \text{ nMoles}/\mu\text{L}$**

**$100 \mu\text{M} = 100 \mu\text{Moles/L}$**

$= 100 \mu\text{Moles} / 1,000 \text{ mL}$

or

$100 \mu\text{Moles} / 10^3 \text{ mL}$

$= 100,000 \text{ nMoles} / 1,000 \text{ mL}$

or

$100 \times 10^3 \text{ nMoles} / 10^3 \text{ mL}$

$= 100 \text{ nMoles} / \text{ mL}$

$= 100 \text{ nMoles} / 1,000 \mu\text{L}$

or

$100 \text{ nMoles} / 10^3 \mu\text{L}$

**$= 0.1 \text{ nMoles}/\mu\text{L} = 100 \mu\text{M}$**



## 5' - LC705-AAgAgTTCggCATCAAggAgCATgg--PH

### Amount / Concentration of product in 1 ml

Synthesis scale and purification :

Modifications :

Number of bases :

Wobble bases and GC content :

3,0 nmol / 3,0  $\mu$ M

Synthesis: 0.00  $\mu$ mol Purification: HPLC Condition: 5LC 3 nmol ly no

A: 8 G: 9 C: 4 T: 4 total 25

Wobbel: 0 Mod.: 0 GC-content 52.0 %

### Chemical properties and constant factors of the product :

Molar extinction coefficient  $\epsilon$

291490 l / mol cm

Molecular weight ammonium salt NH<sub>4</sub>

8188,9 g / mol

Molecular weight free acid :

7780,2 g / mol

Picomoles per OD<sub>260</sub>

3430,6 pmol / OD

Micrograms per OD<sub>260</sub>

28,1  $\mu$ g / OD

### Delivered amount (per vial when delivered in aliquots)

Amount in optical units OD<sub>260</sub>

0,9 OD

Molar amount :

3,0 nmol

Amount in  $\mu$ g mass units :

24,7  $\mu$ g

### Molar concentration when delivered in 1 ml solution :

3,0  $\mu$ M (pmol /  $\mu$ l)

20  $\mu$ M ( 20 pmol/ $\mu$ l) requires a volume of :

151  $\mu$ l

50  $\mu$ M ( 50 pmol/ $\mu$ l) requires a volume of :

60  $\mu$ l (To prepare stock solutions of

100  $\mu$ M ( 100 pmol/ $\mu$ l) requires a volume of :

30  $\mu$ l different concentration)

### Mass concentration (for hybridization) :

Concentration, when dissolved in 1 ml :

0,025  $\mu$ g /  $\mu$ l

Dilution when preparing a solution with 0,5  $\mu$ g/ml :

1 : 5 dilution factor from a 1 ml solution

To prepare a 0,1  $\mu$ g /  $\mu$ l solution dissolve the product in :

247  $\mu$ l

### Melting point, thermodynamic approach (TIB MOLBIOL)

64,9  $^{\circ}$ C

Melting point in the case of a single mutation (-3,5  $^{\circ}$ C)

61,4  $^{\circ}$ C

Suggested PCR annealing temperature ( $\leq$  72 $^{\circ}$ C) :

72,1  $^{\circ}$ C

Melting point GC/AT rule (A/T = 2 $^{\circ}$ C, G/C = 4 $^{\circ}$ C )

76,0  $^{\circ}$ C

Melting point G/C-content rule

61,8  $^{\circ}$ C

### Thermodynamic parameter for the double stranded hybrid :

$\Delta G / \Delta H / \Delta S$  -189.1 / -797.0 / -2040.2 kJ / mol

### Code for degenerated base positions (wobble positions IUB Code)

S = G/C Y = C/T M = A/C H = A/C/T D = A/G/T N = A/C/G/T X = Modif.  
W = A/T R = A/G K = G/T B = C/G/T V = A/C/G I = Inosin s = Thioate

5'- GAC GCA AAA ACA AAA GCA AA -3'

**Properties**

*T<sub>m</sub>* (50mM NaCl): 50.9 °C  
 GC Content: 35. %  
 Molecular Weight: 6,154.1  
 nmoles/OD260: 4.6  
 ug/OD260: 28.2  
 Ext. Coefficient: 218,600 L/(mole·cm)

**Amount Of Oligo**

34.0 = 155.5 = 0.96  
 OD 260 nMoles mg

**Shipped To**

NYS DEPARTMENT OF HEALTH WADSWC  
 ESP -WCLR BIGGS LABS #E224  
 ALBANY, NY 12237  
 USA  
 5184743853  
 Customer No. ████████ PO No. ████████

**Secondary Structure Calculations**

Lowest folding free energy (kcal/mole): -0.31 at 25 °C  
 Strongest Folding *T<sub>m</sub>*: 29.0 °C

**Oligo Base Types**

Oligo Base Types	Quantity
A Bases	20

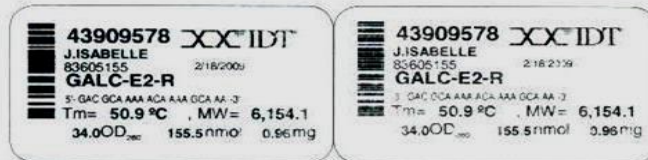
**Disclaimer**

See on reverse page notes (I) (II) & (III) for usage, label license, and product warranties

**Modifications And Services**

Modifications And Services	Quantity
Standard Desalting	1

Mfg. ID ████████ Labels - Peel Here

**I N S T R U C T I O N S**

- Lyophilized contents may appear as either a translucent film or a white powder. This variance does not affect the quality of the oligo

- Please centrifuge tubes prior to opening. Some of the product may have been dislodged during shipping.



**Concept: 0.1 nMoles/ $\mu$ L = 100  $\mu$ M**

**Primer** reported Amount: **155.5 nMoles**

If you want a **Stock Solution** of 100  $\mu$ M, resuspend oligo in 1555  $\mu$ L of ddH<sub>2</sub>O or TE buffer:

$$\bullet 155.5 \text{ nMoles} / 1550 \mu\text{L} = \mathbf{0.1 \text{ nMoles}/\mu\text{L} = 100 \mu\text{M}}$$

If you want a **Working Solution** of 10  $\mu$ M make a 1:10 dilution from the stock:

- Add 1.0  $\mu$ L from Stock (100  $\mu$ M) to 9.0 of ddH<sub>2</sub>O or TE buffer (1:10).

$$\bullet 100 \mu\text{M} \times 1 \mu\text{L} / 10 \text{ uL} = \mathbf{10 \mu\text{M}}$$

**Concentration** in your final PCR mix:  $C_1V_1 = C_2V_2$

$$\bullet 10 \mu\text{M} \times V_1 = 0.3 \mu\text{M} \times 20 \mu\text{L} \longrightarrow V_1 = \frac{0.3 \mu\text{M} \times 20 \mu\text{L}}{10 \mu\text{M}} = 0.6 \mu\text{L}$$

**Primer** reported Amount: 155.5 nMoles = **0.96 mg** = 960  $\mu\text{g}$

If you want a **Stock Solution** of 1  $\mu\text{g}/\mu\text{L}$ , resuspend oligo in 960  $\mu\text{L}$  of ddH<sub>2</sub>O or TE buffer:

$$\bullet 960 \mu\text{g} / 960 \mu\text{L} = \mathbf{1.0 \mu\text{g}/\mu\text{L}} = \mathbf{1000 \text{ ng}/\mu\text{L}}$$

If you want a **Working Solution** of 100  $\text{ng}/\mu\text{L}$  make a 1:10 dilution from the stock:

- Add 1.0  $\mu\text{L}$  from Stock (1  $\mu\text{g}/\mu\text{L}$ ) to 9.0  $\mu\text{L}$  of ddH<sub>2</sub>O or TE buffer (1:10).

$$\bullet 1000 \text{ ng} \times 1 \mu\text{L} / 10 \mu\text{L} = \mathbf{100 \text{ ng}/\mu\text{L}}$$

**Concentration** in your final PCR mix:  $C_1V_1 = C_2V_2$

$$\bullet 100 \text{ ng}/\mu\text{L} \times V_1 = 3 \text{ ng}/\mu\text{L} \times 20 \mu\text{L} \quad \longrightarrow \quad V_1 = \frac{3 \text{ ng}/\mu\text{L} \times 20 \mu\text{L}}{100 \text{ ng}/\mu\text{L}} = 0.6 \mu\text{L}$$

<http://www.idtdna.com/Calc/resuspension/>



Prepare Stock Solution and Working solution for primers forward and reverse to amplify exon 5 of the ABCD1 gene.

You have received:

ABCD1-E5-F1: 183.5 nMoles

ABCD1-E5-R1: 354.8 nMoles

Once prepared your primers do the calculations needed to run a PCR reaction using a 0.25  $\mu$ M concentration for each primer in a 20  $\mu$ L total volume PCR.

